

CELLGENIC™

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Gly-Glu-Pro-Pro-Pro-Gly-Lys-Pro-Ala-Asp-Asp-Ala-Gly-Leu-Val

Gly-His-Lys (Cu^{2+} complex)

Ala-Glu-Ile-Gln-Ala-Ala-Gly-Ala-Asp-Ala-Gly-Gly-Ala-Ala-Ala-Gly-Ala-Ala-Ala-Gly-Ala-Ala-Ala-Gly-Ala-Ala-Ala-Gly-Ala-Ala-A
Ala-Gly-Ala-Ala-Ala-Gly-Ala-Ala-Ala-Gly-Ala-Ala-Ala-Gly-Ala-Ala-Ala

Sample Name	Batch Number	BBG70	Date Published	2025-12-6
Glow Stack	BPC-157 - 10 mg			
	GHK-Cu - 50 mg			
	TB-500 - 10 mg			

The product “Glow Stack” (Batch BBG70) is a multi-component lyophilized peptide formulation containing BPC-157 (10 mg), GHK-Cu (50 mg), and TB-500 (10 mg) per vial. For identity and assay testing, the sample was prepared in accordance with the laboratory method and analyzed under validated chromatographic conditions. Each component was evaluated against its corresponding certified reference standard for identity confirmation and quantitative determination. Purity values reported correspond to primary peak area normalization for each target component under the stated analytical conditions.

Analysis of Peptide Identity, Content and Purity	Result	Unit	Uncertainty	Reporting Limit
GHK-CU Identification by spectrum Peptide Screening	405		[± 20]	
GHK-CU Assay Peptide Screening	50.25	mg	[± 0.03]	
GHK-CU Identification by RT Peptide Screening	4.32		[± 0.05]	
GHK-CU Purity Peptide Screening	96.6	%	[± 0.5]	
Analysis of Peptide Identity, Content and Purity	Result	Unit	Uncertainty	Reporting Limit
BPC-157 Identification by spectrum Peptide Screening	993		[± 20]	
BPC-157 Assay Peptide Screening	10.25	mg	[± 0.03]	
BPC-157 Identification by RT Peptide Screening	11.05		[± 0.05]	
BPC-157 Purity Peptide Screening	97.6	%	[± 0.5]	

Analysis of Peptide Identity, Content and Purity	Result	Unit	Uncertainty	Reporting Limit
TB 500 Identification by spectrum Peptide Screening	4963		[± 20]	
TB 500 Assay Peptide Screening	10.25	mg	[± 0.03]	
TB 500 Identification by RT Peptide Screening	18.90		[± 0.08]	
TB 500 Purity Peptide Screening	97.6	%	[± 0.5]	

Bioburden	Result	Unit	Uncertainty	Reporting Limit
Total Aerobic Microbial Count USP <61> Plate Count Method	Not detected	CFU/g		>= 1000
Total Yeast and Mold Count USP <61> Plate Count Method	Not detected	CFU/g		>= 100

Endotoxin Analysis	Result	Unit	Uncertainty	Reporting Limit
Bacterial Endotoxin Final Combined Sample - Glow Stack USP<85> Bacterial Endotoxin Chromogenic Test	< 0.001	EU/mg		> 0.5

Endotoxin testing was performed on a composite preparation representative of Batch BBG70 (Glow Stack), following USP <85> chromogenic methodology.

Heavy Metals	Result	Unit	Uncertainty	Reporting Limit
Arsenic Elemental Impurities Screening	Not detected	ppm		>= 1.5
Cadmium Elemental Impurities Screening	Not detected	ppm		>= 0.5
Cobalt Elemental Impurities Screening	Not detected	ppm		>= 25

Heavy Metals	Result	Unit	Uncertainty	Reporting Limit
Lead Elemental Impurities Screening	Not detected	ppm		>= 1.5
Nickel Elemental Impurities Screening	Not detected	ppm		>= 25
Quicksilver Elemental Impurities Screening	Not detected	ppm		>= 1.5
Vanadium Elemental Impurities Screening	Not detected	ppm		>= 25

Method Specification

Determination of identity, content and purity

1. Content Assessment

1.1. Instrumentation

Module	Name	Serial Number
System Controller	Shimadzu SCL-10ADvp	C21014112659
Degassing Unit	Shimadzu DGU-14A	NA
Pump A	Shimadzu LC-10ADvp	C20964130075
Pump B	Shimadzu LC-10ADvp	C20953770781
Autosampler	Shimadzu SIL-10ADvp	C21054109114
Colum Thermostat	Shimadzu CTO-10ACvp	C21033770144
Detector	Shimadzu SPD-10ADvp	C20994233588

1.2. Chromatographic conditions

Chromatographic conditions	
Eluent A	0.1% TFA in Water (HPLC, Gradient Grade)
Eluent B	0.1% TFA in Acetonitrile (HPLC, Gradient Grade)
Flow rate	0.4 mL/min
Program	Gradient elution
Injection volume	0.5 µL
Colum Temperature	60°C
Column	Phenomenex Biozen Peptide Polar C18, 150x2.1mm 3µm
Detection wavelength	214nm

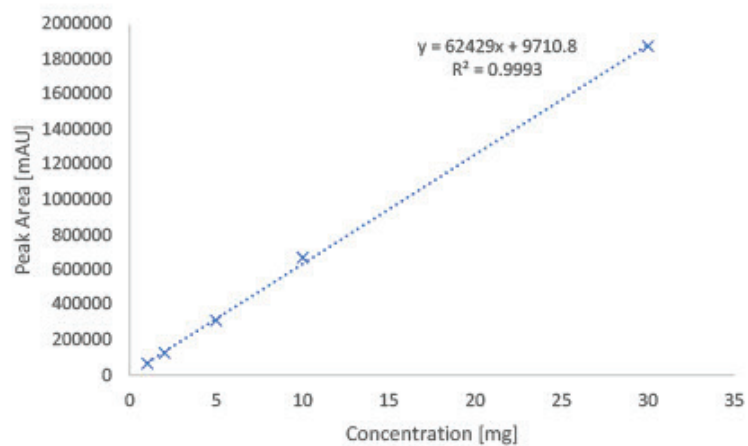
Gradient Program		
Time [min]	A [%]	B [%]
1	95	5
19	40	60
20	5	95
24	5	95
25	95	5
33	end	

1.3. Sample preparation

Whole amount of container was dissolved in 2mL of water (HPLC, Gradient Grade). Aliquote part of 1 mL was dispensed into HPLC vial for analysis.

1.4. Calibration curve

Calibration curve detail	
Quantitative method	External Standard
Calibration Type	Linear
Number of calibration points	5
Force through Zero	Disabled
Weighting Method	None



2. Purity assessment

2.1 Instrumentation

Module	Name	Serial Number
System Controller	Shimadzu SCL-10ADvp	C21014112659
Degassing Unit	Shimadzu DGU-14A	NA
Pump A	Shimadzu LC-10ADvp	C20964130075
Pump B	Shimadzu LC-10ADvp	C20953770781
Autosampler	Shimadzu SIL-10ADvp	C21054109114
Column Thermostat	Shimadzu CTO-10ACvp	C21033770144
Detector	Shimadzu SPD-10ADvp	C20994233588

2.2 Chromatographic conditions

Chromatographic conditions	
Eluent A	0.1% TFA in Water (HPLC, Gradient Grade)
Eluent B	0.1% TFA in Acetonitrile (HPLC, Gradient Grade)
Flow rate	0.4 mL/min
Program	Gradient elution
Injection volume	0.5 µL
Column Temperature	60°C
Column	Phenomenex Biozen Peptide Polar C18, 150x2.1mm 3µm
Detection wavelength	214nm

Gradient Program		
Time [min]	A [%]	B [%]
1	95	5
19	40	60
20	5	95
24	5	95
25	95	5
33	end	

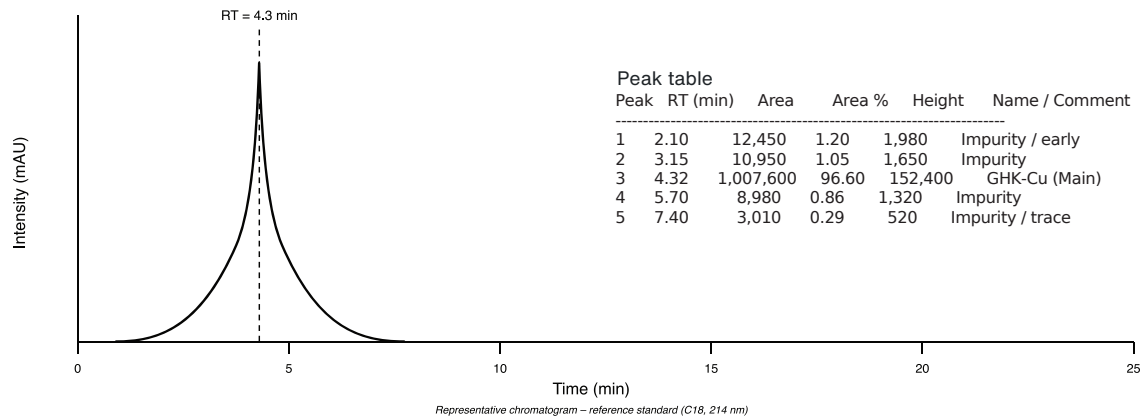
1.5. Sample preparation

Whole amount of container was dissolved in 2mL of water (HPLC, Gradient Grade). Aliquote part of 1 mL was dispensed into HPLC vial for analysis.

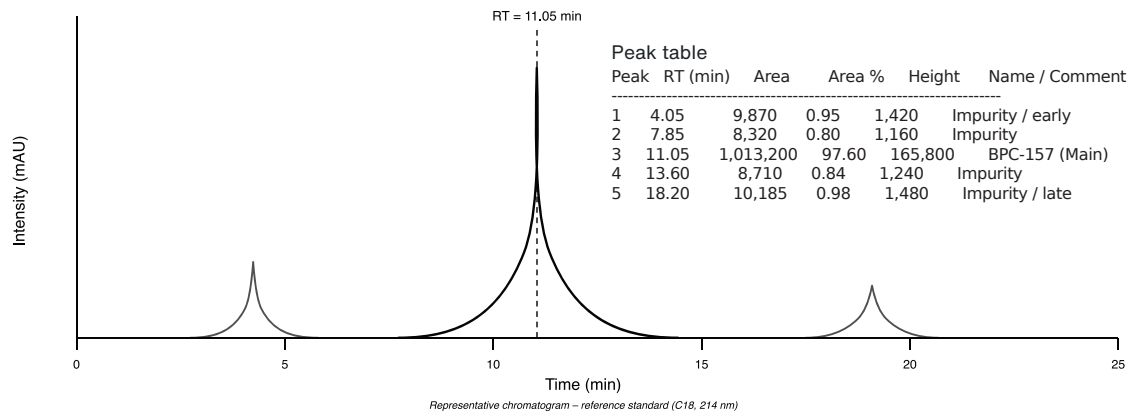
1.6. Purity assesment

Purity of compound assesed by area normalization method, comparing area of each peak to sum of area of all peaks detected at wavelenght of 214 nm.

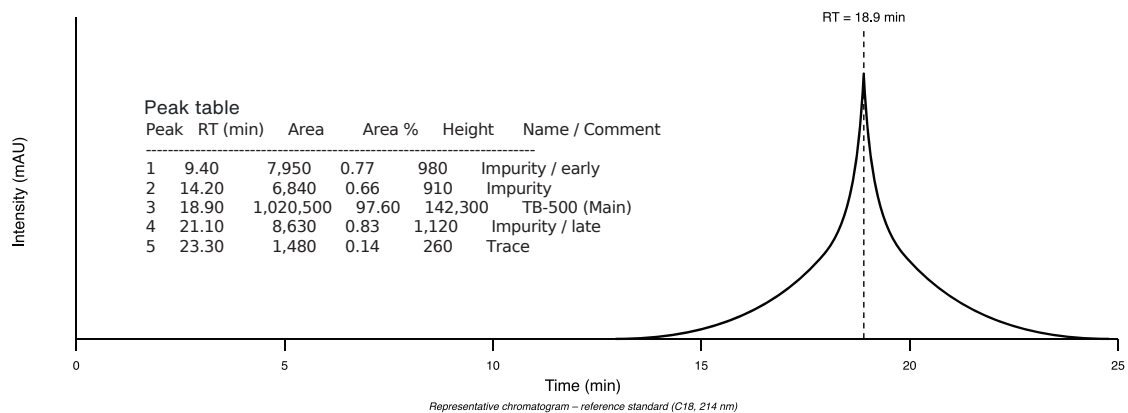
GHK-Cu – Representative HPLC Chromatogram



BPC-157 – Representative HPLC Chromatogram



TB-500 – Representative HPLC Chromatogram



Peak tables are representative for reference standards analyzed under comparable HPLC conditions.

1. Instrumentation and chemicals

1.1. Instruments used

- Sterile Syringe 2mL Luer
- Sterile needles
- Ready made PCA Plate ROTI Aquatest
- Ready made Sab4 Plate ROTI Aquatest

1.2. Chemicals

Sterile physiological solution (0.9% NaCl)

2. Sample preparation and inoculation

2.1 Sample preparation

1. Fresh sterile needle and syringe was used for measuring exactly 2 mL of sterile physiological solution.
2. Needle was changed and by new needle rubber top of peptide container was penetrated and 2 mL of sterile physiological solution was dispensed.
3. Content of container was completely dissolved and left for 5 minutes to settle potentially created bubbles.
4. This procedure is repeated for two vials.

2.2 Total Aerobic microbial count inoculation and cultivation

1. By sterile needle 1 mL of solution was filled into the sterile syringe.
2. Needle was placed above the flame for few seconds to sterilize.
3. Consequently 1 mL of solution was poured into the ready to use sterile petri dish filled with PCA agar and petri dish was closed.
4. Process was repeated for two petri dishes.
5. With sterile needle, 1 mL of sterile physiological solution was filled into the sterile needle and was inoculated onto one sterile petri dish filled with PCA agar as negative control sample.
6. Samples and negative control sample were placed in incubator at temperature 37°C for 120h.

2.3 Total Yeast and Mold count inoculation and cultivation

1. By sterile needle 1 mL of solution was filled into the sterile syringe.
2. Needle was placed above the flame for few seconds to sterilize.
3. Consequently 1 mL of solution was poured into the ready to use sterile petri dish filled with Sab4 agar and petri dish was closed.
4. Proces was repeated for two petri dishes.
5. With sterile needle, 1 mL of sterile physiological solution was filled into the sterile needle and was inoculated onto one sterile petri dish filled with Sab4 agar as negative control sample.
6. Samples and negative control sample were placed in incubator at temperature 25°C for 72h.

3. Evaluation of results

After incubation time, colonies are counted as cfu (colonies forming units) and result per 1g of sample is determined as:

$$CFU_{avg} = \frac{\sum CFU_n}{n}$$

CFU_{avg} = average CFU counted form n inoculations

CFU_n = CFU counted per inoculation

n = number of inoculations

$$CFU \text{ per gram} = \frac{CFU_{avg}}{m_s} * DF$$

CFU_{avg} = Average CFU counted from n inoculations

m_s = mass of sample (mg)

DF = Dilution factor

If negative control sample is evaluated as positive, process have to be repeated due to possible contamination in the process of inoculation or incubation.

1. Chromgenic LAL Assay Determination of Bacterial Endotoxin content of sample

1.1. Instrumentation

- Pipette set 1-1000 µL
- Thermostatically controlled water bath
- UV VIS spectrometer (Shimadzu UV-1601)
- GenScript ToxinSensor Chromgenic LAL Endotoxin Assay kit

1.2. Chemicals

- LAL Reagent water (endotoxin free)
- Limulus Amoebocyte Lysate
- LAL Substrate
- Color Stabilizer #1
- Color Stabilizer #2
- Color Stabilizer #3
- 35% HCl (p.a.)

1.3. Sample preparation

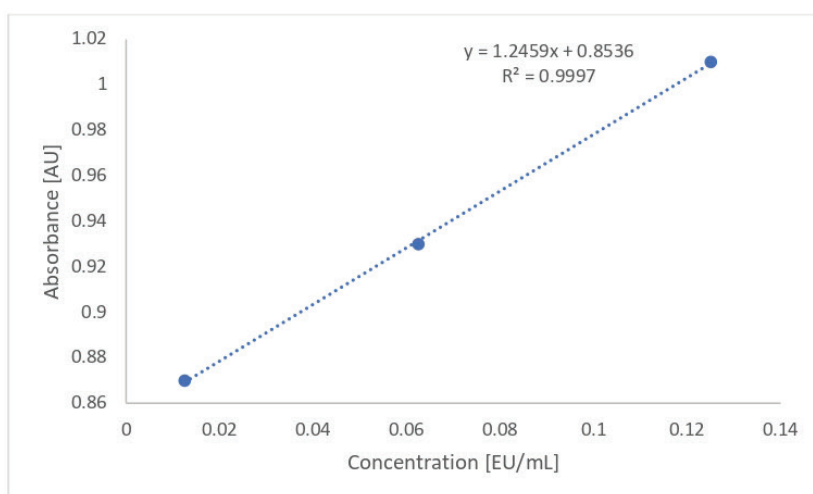
1. Sample container was weighed prior to dissolution and measured weight was marked.
2. Sample was completely dissolved in its container by 2 mL of LAL Reagent water.
3. 100 µL of the sample was aliquoted for analysis.
4. After analysis container was emptied and dried.
5. Dry mass of container was measured and exact weight of dissolved content was determined as:

$$m_{dc} = m_{sample} - m_{container}$$

1.5. Measurement procedure

	Standards	Samples	Blank
Standards (mL)	0.1	-	-
Samples (mL)	-	0.1	-
LAL Reagent Water (mL)	-	-	0.1
LAL Solution (mL)	0.1	0.1	0.1
Mix well and incubate at 37°C for 27 min			
Substrate solution (mL)	0.1	0.1	0.1
Mix well and incubate at 37°C for 6 min			
Color Stabilizer #1 solution	0.5	0.5	0.5
Color Stabilizer #2 solution	0.5	0.5	0.5
Color Stabilizer #3 solution	0.5	0.5	0.5
Mix well and read the absorbance at 545nm			

1.6. Calibration curve



1.7. Calculation of endotoxin content

Endotoxin content of the sample was calculated from the calibration curve as:

$$Endotox[EU/mg] = \frac{\left(\frac{ABS_{sample}}{S_{calib}}\right) * 20}{m_{sample}}$$

ABS_{sample} = Measured absorbance of sample

S_{calib} = Slope of calibration curve

m_{sample} = real measured mass of sample

20 = dilution factor of measured sample

Analysis results relate only to the samples tested.

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Inspecting QA/QC Manager

